

ANALOGS OF DEAMINO CARBA OXYTOCIN WITH INHIBITORY PROPERTIES; SYNTHESIS AND BIOLOGICAL ACTIVITIES*

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Solid phase methodology on polyamide-kieselguhr resin was used for the synthesis of six analogs of deamino carba-1 or carba-6 oxytocin with non-coded amino acids in position 2, threonine in position 4, ornithine in position 8 and without glycine in position 9. The following analogs were prepared: des-Gly⁹-[L-Phe(*p*-Et)², Thr⁴, Orn⁸]deamino-carba-1-oxytocin (*I*), des-Gly⁹-[D-Phe(*p*-Et)², Thr⁴, Orn⁸]deamino-carba-1-oxytocin (*II*), des-Gly⁹-[D-Tyr(Et)², Thr⁴, Orn⁸]deamino-carba-1-oxytocin (*III*), des-Gly⁹-[L-Phe(*p*-Et)², Thr⁴, Orn⁸]deamino-carba-6-oxytocin (*IV*), des-Gly⁹-[D-Phe(*p*-Et)², Thr⁴, Orn⁸]deamino-carba-6-oxytocin (*V*), and des-Gly⁹-[D-Tyr(Et)², Thr⁴, Orn⁸]deamino-carba-6-oxytocin (*VI*). All the analogs were found to be strong uterotonic and pressor inhibitors. The highest potency in the uterotonic inhibitory test was exhibited by analog *II* ($pA_2 = 8.3$) and the strongest pressor inhibitor was compound *I* ($pA_2 = 7.5$).

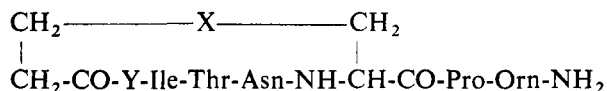
Highly potent and selectively acting uterotonic inhibitor might find its application in human medicine especially if it would have a prolonged time course of action. Structure-activity studies in the field of neurohypophyseal hormone analogs** (for recent reviews see refs^{1,2}) have led to discovery of several structural features yielding analogs with inhibitory properties. It is well documented that shortening of the peptide chain at the carboxy terminal part of the molecule destroys agonistic activities, but does not influence the inhibitory potency of the analog^{3,4}. Ornithine in position 8 represents a very important modification for the uterotonic inhibitory activity^{5,6}. On the other hand, the effects of threonine in position 4, deamination in position 1 and carba substitution of the disulfide bridge on the inhibitory activity

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** All the chiral amino acids, mentioned in this work, are of the L-series, if not specified otherwise. The nomenclature and symbols of the amino acids and peptides obey the published recommendations (Eur. J. Biochem. 138, 9 (1984)). Phe(*o*-Et) denotes *p*-ethylphenylalanine and Tyr(Et) the O-ethyltyrosine moiety.

are not clearly defined. All these modifications increase the agonistic activities and therefore, they are supposed to enhance also the affinity to the appropriate receptor. However, for example the combination of deaminopenicillamine in position 1 (normally leading to inhibitor) with carba substitution resulted in a quite potent uterotonic agonist which was transformed to a potent antagonist only by another modification of the tyrosine side chain⁷. Substitution of tyrosine hydroxy group, either by alkoxy or alkyl group, usually gives inhibitors or at least partial agonists (see e.g. refs^{8,9}), as well as the introduction of amino acids of D-configuration into position 2 (ref.¹⁰). For recent review of structure-activity studies in the field of neurohypophyseal hormone inhibitors see ref.¹¹. We decided to combine the structural features stated above to study their cooperativity for production of a potent uterotonic inhibitor.

All the synthesized analogs are of the deamino series (aminogroup on amino acid moiety in position 1 is replaced with hydrogen atom), without glycine in position 9 (des-glycine⁹-analogs); glutamine in position 4 is replaced by threonine and leucine in position 8 by ornithine. The disulfide bridge is modified by the so called carba substitution. In the first three analogs, the sulfur atom in position one is replaced with a methylene group (carba-1 analogs), in the last three analogs position six is modified (carba-6 analogs). In position 2, there are non-coded amino acids L- and D-*p*-ethylphenylalanine and D-O-ethyltyrosine. The following analogs were prepared: des-Gly⁹-[L-Phe(*p*-Et)², Thr⁴, Orn⁸]deamino-carba-1-oxytocin (I), des-Gly⁹-[D-Phe(*p*-Et)², Thr⁴, Orn⁸]deamino-carba-1-oxytocin (II), des-Gly⁹-[D-Tyr(Et)², Thr⁴, Orn⁸]deamino-carba-1-oxytocin (III), des-Gly⁹-[L-Phe(*p*-Et)², Thr⁴, Orn⁸]deamino-carba-6-oxytocin (IV), des-Gly⁹-[D-Phe(*p*-Et)², Thr⁴, Orn⁸]deamino-carba-6-oxytocin (V), and des-Gly⁹-[D-Tyr(Et)², Thr⁴, Orn⁸]deamino-carba-6-oxytocin (VI).



- I, X = CH₂-S; Y = L-Phe(*p*-Et)
 II, X = CH₂-S; Y = D-Phe(*p*-Et)
 III, X = CH₂-S; Y = D-Tyr(Et)
 IV, X = S-CH₂; Y = L-Phe(*p*-Et)
 V, X = S-CH₂; Y = D-Phe(*p*-Et)
 VI, X = S-CH₂; Y = D-Tyr(Et)

Pentafluorophenyl ester of N²-fluorenyloxycarbonyl-S-(3-tert-butoxycarbonylpropyl)cysteine and pentafluorophenyl ester of N²-fluorenyloxycarbonyl-S-(2-tert-butoxycarbonylethyl)homocysteine served as key intermediary products for the synthesis of carba analogs. These intermediates were prepared from S-(3-tert-butoxycarbonylpropyl)cysteine and S-(2-tert-butoxycarbonylethyl)homocysteine^{12,13}

via dicyclohexylammonium salts of the corresponding N^{α} -fluorenylmethyloxycarbonyl derivatives. S-(3-Tert-butoxycarbonylpropyl)cysteine was obtained by alkylation of cysteine by tert-butyl ester of γ -bromobutyric acid. S-(2-Tert-butoxycarbonyl-ethyl)homocysteine was prepared by reduction of homocysteine with sodium in liquid ammonia and subsequent alkylation of the homocysteine with tert-butyl acrylate.

Syntheses of all the six analogs were performed by solid phase technique on the polyamide-kieselguhr resin. As the α -amino group protection we have used fluorenylmethyloxycarbonyl group, only for the aminoterminal amino acid tert-butoxycarbonyl group was used. For the side chain protection we have used benzyloxycarbonyl (Orn) or tert-butyl (Thr) group. Protected amino acids were coupled by the pentafluorophenyl active ester method in dimethylformamide. Acid-labile side chain protecting groups (Boc and Bu^t) were cleaved with trifluoroacetic acid before the cleavage of the peptide from the resin by liquid ammonia in methanol. The cyclization was carried out in dimethylformamide by diphenylphosphorylazide with dipotassium hydrogen phosphate¹⁴ and stable side chain protecting group (Z on ornithine) was cleaved by liquid hydrogen fluoride with anisole. Cyclic analogs were isolated and purified by HPLC. In syntheses of the analogs with *p*-ethylphenylalanine we took advantage of fact that diastereoisomeric peptides can be easily separated by reversed phase chromatography^{10,15,16}. Preparation of these analogs was carried out with the mixture of L- and D-amino acids and the corresponding diastereoisomeric analogs were separated by HPLC at the end of the synthesis. With the knowledge that with sufficient excess of L and D components only one diastereoisomeric derivative¹⁷ is predominantly formed, only 1:1 equivalents of tert-butoxycarbonyl-D,L-*p*-ethylphenylalanine were used in this case.

Biological activities of the analogs are given in Table I. Since activities of the

TABLE I

Inhibitory activities (rat) of oxytocin analogs (pA_2 values). Values in parentheses were determined independently in Malmö laboratory

Compound	Uterotonic		Duration ^a min	Pressor
	in vitro	in vivo		
I	8.05 $\pm$ 0.20 (8.20 $\pm$ 0.07)	7.42 $\pm$ 0.19 (6.84 $\pm$ 0.24)	51 $\pm$ 8	7.55
II	8.32 $\pm$ 0.13 (8.39 $\pm$ 0.21)	7.44 $\pm$ 0.25 (7.73 $\pm$ 0.16)	81 $\pm$ 14	6.83 $\pm$ 0.18
III	8.15 $\pm$ 0.10 (8.15 $\pm$ 0.14)	6.75 $\pm$ 0.08 (6.92 $\pm$ 0.17)	60 $\pm$ 9	6.56
IV	8.43 $\pm$ 0.25	7.45 $\pm$ 0.24 (7.45 $\pm$ 0.15)	87 $\pm$ 11	7.43 $\pm$ 0.21
V	8.50 $\pm$ 0.18 (8.35 $\pm$ 0.05)	7.40 $\pm$ 0.13 (7.51 $\pm$ 0.07)	140 $\pm$ 19	6.42 $\pm$ 0.09
VI	8.75 $\pm$ 0.20 (8.48 $\pm$ 0.07)	7.70 $\pm$ 0.25 (7.70 $\pm$ 0.21)	99 $\pm$ 19	6.9 (6.12 $\pm$ 0.04)

^a Time after which the response to standard dose of oxytocin returned to 75% of its starting value; see ref.⁹.

described compounds were found to be quite promising, their potencies were evaluated both in Prague and Malmö laboratories. This independent evaluation has shown, that the inhibitory activities determined at different places can be well correlated (see Table I) and that highly potent uterotonic inhibitors resulted from the substitutions performed on the oxytocin molecule. The highest pA_2 value (8.75) was achieved with analog VI, containing O-ethyltyrosine of D-configuration in position 2. This modification is slightly more efficient in producing an inhibitor than D-*p*-ethylphenylalanine ($pA_2 = 8.5$) which was found by several authors⁹ to be the optimum substitution for achieving inhibitory properties of the analog. Comparison of carba-1 and carba-6 substitutions of the disulfide bridge (I and IV, II and V, III and VI), favours the carba-6 modification. Evaluation of the in vivo inhibitory potency of these compounds showed that there are no substantial differences in the character of the response between the analogs containing L- or D-*p*-ethylphenylalanine in position 2. However, carba-6 analogs exhibit substantially more prolonged inhibitory action than carba-1 analogs. This may be explained by smaller influence of oxidation (which may play a role as one of the degradation processes) on the activity of carba-6 analogs in comparison to carba-1 ones, as was described in several cases¹⁸. The action of analogs containing D-Phe(Et) is by about 40% more prolonged than of the corresponding analogs with D-Tyr(Et) in position 2. This may be caused by cleavage of the *p*-substituent in the tyrosine derivative. Analog V is by far the most prolonged uterotonic inhibitor we have ever tested.

The most potent pressor inhibitor was the analog IV containing *p*-ethylphenylalanine of L-configuration ($pA_2 = 7.6$) in position 2. The position of the sulfur replacement by CH₂ apparently has not a dramatic influence on the pressor inhibitory activity.

We may conclude that the compounds synthesized as described in this paper, belong to the family of uterotonic antagonists with prolonged action. They may have a potential clinical value in preventing oxytocin receptor-dependent uterine hyperactivity in premature labour.

EXPERIMENTAL

Thin-layer chromatography (TLC) was carried out on silica gel coated plates (Silufol, Kavalier, Czechoslovakia) in the following systems: 2-butanol-98% formic acid-water (10 : 3 : 8) (S1), 2-butanol-25% ammonia-water (85 : 7.5 : 7.5) (S2), 1-butanol-acetic acid-water (4 : 1 : 1) (S3), 1-butanol-acetic acid-pyridine-water (15 : 3 : 10 : 6) (S4). Spots in TLC were detected by the chlorination method. Samples for amino acid analysis were analyzed on an Amino acid analyser LKB 4150. Optical rotations were determined on a Perkin-Elmer instrument type 141 MCA (Norwalk, U.S.A.) at 20°C. Fast atom bombardment mass spectra were obtained on a ZAB-EQ spectrometer (VG Analytical Ltd., Manchester) with xenon at 8 kV as the bombarding gas. Analytical HPLC was carried out on an SP-8800 instrument equipped with a Spectroflow 757 detector (Applied Biosystems) and SP-4290 integrator (SP from Spectra Physics, Santa Clara,

U.S.A.) and on a Waters μ BONDAPAK C18 column. Preparative liquid chromatography was carried out on SepTech ST/Lab 800C instrument with Macrobore Annular Expansion column A/E (25×7.5 cm) filled with Kromosil C18 ($10 \mu\text{m}$).

**N^α-Fluorenylmethyloxycarbonyl-N^δ-benzyloxycarbonylornithine
Dicyclohexylammonium Salt**

N^δ-Benzyloxycarbonylornithine¹⁹ (7.3 g; 20 mmol) was suspended in the mixture of water (50 ml) and acetonitrile (100 ml) and diisopropylethylamine (6.8 ml) and Fmoc-ONSu (7.4 g; 22 mmol) were added. After stirring for 2 h at room temperature acetonitrile was evaporated, the residue was acidified with 1M HCl and the product was extracted into ethyl acetate. Extract was washed with water, dried with sodium sulfate, filtered and evaporated. The residue was dissolved in ethyl acetate and after addition of dicyclohexylamine (4 ml) the mixture was diluted with hexane. After several hours in the refrigerator, the crystals were filtered and washed with hexane. Yield 11.5 g (82%); m.p. $220-221^\circ\text{C}$; $[\alpha]_D +7.5^\circ$ (c 0.52, ethanol). For $\text{C}_{40}\text{H}_{51}\text{N}_3\text{O}_6$ (669.9) calculated: 71.72% C, 7.67% H, 6.27% N; found: 71.95% C, 7.57% H, 6.31% N.

S-(3-Tert-butoxycarbonylpropyl)cysteine

Cystine (10 g) was dissolved in liquid ammonia (500 ml) and sodium was added until blue color persisted for 5 min. The solution was decolorized by addition of ammonium chloride, and tert-butyl ester of γ -bromobutyric acid (28 g) was added under stirring. Solution was evaporated in vacuo of water aspirator, the lyophilizate was dissolved in water (250 ml) and extracted with ethyl acetate. The aqueous layer was cooled to 0°C and applied to the column of Dowex 50 (H^+ form, 300 ml). The ion exchanger was washed with water to neutral reaction of the eluate and the product was eluted with 5% aqueous ammonia. Evaporation afforded 18.6 g (86%) of the cysteine derivative. $E_{2.4}^{G/y}$ 0.48 (0.48), $E_{5.7}^{H/s}$ 0.00 (−0.63) (values in parenthesis are for the diacid prepared by trifluoroacetic acid treatment); $[\alpha]_D -12.8^\circ$ (c 1.0, 1M acetic acid). For $\text{C}_{11}\text{H}_{21}\text{NO}_4\text{S}$ (263.4) calculated: 50.17% C, 8.04% H, 5.32% N; found: 49.83% C, 8.25% H, 5.06% N.

The mixture of the S-(3-tert-butoxycarbonylpropyl)cysteine (1.4 g, 5 mmol), water (7.5 ml) and acetonitrile (7.5 ml) was stirred with diisopropylethylamine (0.85 ml) and Fmoc-ONSu (1.87 g, 5.5 mmol) for 30 min. The product was isolated similarly as above. Yield 2.93 g (86%); m.p. $142-143^\circ\text{C}$; $[\alpha]_D +2.8^\circ$ (c 0.36, ethanol). For $\text{C}_{38}\text{H}_{54}\text{N}_2\text{O}_6\text{S}$ (666.9) calculated: 68.44% C, 8.16% H, 4.20% N; found: 68.22% C, 8.02% H, 4.11% N.

Pentafluorophenyl Ester of N^α-Fluorenylmethyloxycarbonyl-S-(3-tert-butoxycarbonylpropyl)cysteine

N^α-Fluorenylmethyloxycarbonyl-S-(3-tert-butoxycarbonylpropyl)cysteine dicyclohexylammonium salt (2.05 g, 3 mmol) was suspended in ethyl acetate and shaken with 0.5M H_2SO_4 ; the organic layer was washed with water, dried by sodium sulphate, filtered and evaporated. To the residue dissolved in ethyl acetate (6 ml), pentafluorophenol (0.61 g) and dicyclohexylcarbodiimide (0.68 g) were added and after 1 h of stirring at room temperature and 2 h standing in the refrigerator, dicyclohexylurea was filtered off, the filtrate was concentrated and diluted with hexane. After several hours in refrigerator, crystalline portion was filtered and washed with hexane. Yield 1.7 g (86%); m.p. $61-62^\circ\text{C}$; $[\alpha]_D -23.5^\circ$ (c 0.2, methanol). For $\text{C}_{32}\text{H}_{30}\text{F}_5\text{NO}_6\text{S}$ (651.7) calculated: 58.98% C, 4.64% H, 2.15% N; found: 58.92% C, 4.68% H, 2.24% N.

S-(2-Tert-butoxycarbonylethyl)homocysteine

Homocystine²⁰ (8.06 g, 30 mmol) was reduced by sodium in liquid ammonia (300 ml). After the solution had been decolorized by ammonium chloride, the solution was lyophilized in the vacuo of a water aspirator. The lyophilizate was dissolved in water (120 ml, pre-bubbled through with nitrogen) at 0°C and pH of the solution was adjusted to 8.1 by 1M HCl. Tert-butyl acrylate (17.4 ml, 120 mmol) was added to this solution during 15 min and the reaction mixture was stirred under nitrogen overnight. Separated product was filtered off, washed with ethanol and ether. Yield 6.75 g (41%) of electrophoretically pure product, which was found identical to the compound obtained in a different manner¹². Aqueous filtrate was acidified by 1M HCl to pH 3, washed with ether and passed over a column of Dowex 50Wx8 (H⁺-form; 350 ml; precooled to 0°C). The column was washed with water, the product was eluted with 10% solution of pyridine and evaporated to dryness. Yield 6.8 g (41%) of the second crop, containing traces of dicarboxylic acid.

N^α-Fluorenylmethyloxycarbonyl-S-(2-tert-butoxycarbonylethyl)homocysteine Dicyclohexylammonium Salt

This product was obtained from S-(2-tert-butoxycarbonylethyl)homocysteine (2.8 g, 10 mmol) using the same procedure as described above. The crude product was isolated from ethyl acetate and ether (5.72 g, 84%) and was recrystallized from 80% aqueous ethanol. Yield 3.55 g (52%); m.p. 198–201°C; $[\alpha]_D + 13.1^\circ$ (c 0.53, ethanol). For C₃₈H₅₄N₂O₆S (666.9) calculated: 68.44% C, 8.16% H, 4.20% N; found: 68.37% C, 8.01% H, 4.36% N. Amino acid derivative released from its dicyclohexylammonium salt was found identical to that described earlier¹³.

Pentafluorophenyl Ester of N^α-Fluorenylmethyloxycarbonyl-S-(2-tert-butoxycarbonylethyl)homocysteine

N^α-Fluorenylmethyloxycarbonyl-S-(3-tert-butoxycarbonylethyl)homocysteine dicyclohexylammonium salt (3.4 g, 5 mmol) was transformed to the active ester as in the case of the cysteine derivative. Yield 2.32 g (70%); m.p. 58–59°C; $[\alpha]_D - 27.8^\circ$ (c 0.50, methanol). For C₃₂H₃₀F₅NO₆S (651.8) calculated: 58.98% C, 4.64% H, 2.15% N; found: 58.97% C, 4.70% H, 2.36% N.

Pentafluorophenyl Ester of N^α-Tert-butoxycarbonyl-D,L-p-ethylphenylalanine

N^α-Tert-butoxycarbonyl-D,L-p-ethylphenylalanine (0.88 g, 3 mmol) was dissolved in ethyl acetate (6 ml) together with pentafluorophenol (0.61 g) and dicyclohexylcarbodiimide (0.68 g), and after 1 h of stirring at room temperature and 2 h standing in refrigerator, the urea was filtered off; the filtrate was concentrated in vacuo and diluted with hexane. Crystals were separated in the refrigerator and washed with hexane. Yield 0.87 g (63%); m.p. 95–96°C. For C₂₂H₂₂F₅NO₄ (459.4) calculated: 57.52% C, 4.83% H, 3.05% N; found: 57.84% C, 4.78% H, 2.79% N.

Pentafluorophenyl Ester of N^α-Tert-butoxycarbonyl-O-ethyl-D-tyrosine

The title compound was prepared from N^α-tert-butoxycarbonyl-O-ethyl-D-tyrosine (0.96 g, 3 mmol) analogously as above. Yield 1.1 g (77%); m.p. 103–104°C; $[\alpha]_D + 16.1^\circ$ (c 0.51, methanol). For C₂₂H₂₂F₅NO₅ (475.5) calculated: 55.58% C, 4.66% H, 2.95% N; found: 55.39% C, 4.61% H, 2.87% N.

Solid Phase Peptide Synthesis

A) Solid-phase peptide synthesis was carried out on MilliGen 9050 PepSynthesizer instrument

with polyamide-kieselguhr resin (type PepSyn KB; 0.09 meq/g). Synthesis was carried out with 2.2 g resin (0.2 mmol) using the flow-through method on a column and with recycling in the case of coupling. The first amino acid was incorporated by means of the symmetric anhydride method. A cycle for incorporation of the amino acid residue into the growing peptide chain consisted of the following steps: 1. cleaving the Fmoc group by washing with 20% piperidine in dimethylformamide for 10 min; 2. washing with dimethylformamide for 12 min; 3. coupling with 4 equivalents of active ester of the Fmoc-protected amino acid derivative in dimethylformamide for 45 min; (in the case of D,L-*p*-ethylphenylalanine derivative, only 1.1 equivalents of active ester of the Boc-protected amino acid derivative in dimethylformamide for 4 h was used and additional coupling with 0.8 equivalents of the amino acid derivative and 0.8 equivalents of N,N-dimethylaminopyridine for another 1 h); 4. washing with dimethylformamide for 8 min. The following protected derivatives were used: Fmoc-Orn(Z)-OH, DCHA, Fmoc-Pro-OPfp, Fmoc-Cys(C₃H₆CO₂Bu^t)-OPfp (for carba-1 analogs) or Fmoc-Hcy(C₂H₄CO₂Bu^t)-OPfp (for carba-6 analogs), Fmoc-Asn-OPfp, Fmoc-Thr-ODhbt, Fmoc-Ile-OPfp and Boc-D,L-Phe(*p*-Et)-OPfp or Boc-D-Tyr(Et)-OPfp. After taking out the resin from the column, Boc and Bu^t protecting groups were cleaved with the mixture of trifluoroacetic acid, dichloromethane and anisole (45 : 45 : 10; twice 15 min). The resin was washed with dichloromethane, methanol, dichloromethane, 5% diisopropylethylamine in dichloromethane and dichloromethane.

B) Alternative synthesis of analog VI was performed on semiautomatic peptide synthesizer using aminomethyl resin (1 g, 1 mmol NH₂/g, Peptide Intermediates, Prague), substituted by *p*-[(R, S)- α -[1-(9H-fluoren-9-yl)-methoxyformamido]-2,4-dimethoxybenzyl]phenoxyacetic acid²¹. A cycle for incorporation of one amino acid residue into the growing peptide chain consisted of the following steps: 1. cleavage of Fmoc group by 20% piperidine in dimethylformamide for 20 min; 2. washing with dimethylformamide (5 $\times$ 2 min); 3. coupling with 3 equivalents of Fmoc amino acid, HOBt and diisopropylcarbodiimide (monitoring by bromophenol blue method²²); 4. washing with dimethylformamide (5 $\times$ 2 min). The following protected derivatives were used: Fmoc-Orn(Z)-OH, Fmoc-Pro-OH, Fmoc-Hcy(C₂H₄CO₂Bu^t)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Thr(Bu^t)-OH, Fmoc-Ile-OH and Boc-D-Tyr(Et)-OH. After taking out the resin from the reactor and proper washing, peptide was deprotected and cleaved from the resin by the mixture of trifluoroacetic acid (95%), anisole (2%), ethanedithiol (2%), and water (1%) (twice 30 min). The product after evaporation was dissolved in 3M acetic acid and lyophilized. The first treatment afforded 0.5 g of product slightly contaminated with the peptide with deprotected ornithine side chain. The second TFA treatment gave another 0.27 g of the product containing substantial amount of side chain deprotected peptide.

Ammonolytic Cleavage of the Peptide from the Resin (A)

The peptide on the resin was left aside overnight at room temperature in a mixture of methanol (50 ml) and liquid ammonia (100 ml). The resin was filtered off, washed with methanol and the solution was evaporated to dryness. The residue was precipitated from a small amount of methanolic ammonia and ether, filtered and washed with ether. Yields, electrophoretic mobilities and HPLC constants are given in Table II.

The Cyclization (B)

To the solution of the heptapeptide (80–130 mg) in dimethylformamide (20–25 ml), cooled to 0°C, K₂HPO₄ (90–140 mg) and diphenylphosphoryl azide (40–70 μ l) were added. Cyclization was followed by HPLC. If most of the starting compound was not consumed by cyclization in 8 h, another DPPA (100 μ l) was added. After 24 h, the starting compound was never detected.

TABLE II
The values for analogs after ammonolysis (A) and cyclization (B)

Compound	A					B Yield
	Yield	<i>k</i>		<i>E</i> _{2.4} ^{Gly}	<i>E</i> _{5.7} ^{His}	
		60% MeOH	50% MeOH			
<i>I</i>	102 mg (48%) ^b	1.20	6.07	0.61	0.11	^a
<i>II</i>	102 mg (48%) ^b	3.03	20.42	0.61	0.11	^a
<i>III</i>	80 mg (38%)	gradient	—	—	—	60 mg
<i>IV</i>	132 mg (62%) ^b	gradient	—	—	—	120 mg ^b
<i>V</i>	132 mg (62%) ^b	gradient	—	—	—	120 mg ^b
<i>VI</i>	130 mg (61%)	gradient	—	—	—	74 mg

^a Values not determined (product was not desalted). ^b Mixture of diastereoisomers.

TABLE III
The analytical values for analogs after HPLC purification

Compound	<i>I</i>	<i>II</i>	<i>III</i>	<i>IV</i>	<i>V</i>	<i>VI</i>
Gradient ^a , min	60	60	90	120	120	120
Yield, mg	8.9	10.4	13.0	15.4	14.6	38.7
Yield, %	10	12	7	17	16	21
<i>k</i> (25% CH ₃ CN)	8.1	14.5	8.1	8.7 ^b	12.3 ^b	5.0
FAB MS	904	904	919	903.5	903.5	919
Amino acid analysis						
Asp	1.1	1.0	1.1	1.0	1.1	1.1
Thr	1.0	1.0	1.0	1.1	1.0	1.1
Asp	1.1	1.0	1.1	1.0	1.1	1.1
Thr	1.0	1.0	1.0	1.1	1.0	1.1
Pro	0.9	0.9	1.0	0.9	0.9	1.0
Cys(C ₃ H ₅ COOH)	0.8	0.9	0.9	—	—	—
Hcy(C ₂ H ₄ COOH)	—	—	—	0.8	0.8	0.7
Ile	1.0	1.0	0.9	1.0	1.0	1.0
Tyr	—	—	0.6	—	—	1.0
Phe(<i>p</i> -Et)	0.9	0.9	—	0.9	0.9	—
Orn	1.0	1.1	1.0	1.1	1.1	1.2

^a See Experimental. ^b Vydac 218TP54 column.

The reaction mixture was filtered, evaporated, the residue was treated with water, left in a refrigerator, and the aqueous layer was separated from the oily residue by decantation. The oil was dissolved in methanol, dried by evaporation with benzene, and treated with ether. The precipitate was filtered and washed with ether. Yields are given in Table II. In another approach the residue after evaporation of dimethylformamide was dissolved in 3M acetic acid and gel-filtered on Sephadex G-15.

Cleavage of the Benzyloxycarbonyl Group from Ornithine

The cyclic peptide was treated with liquid hydrogen fluoride (5 ml, 10 min, 20°C) in the presence of anisole (0.5 ml). The deprotected peptide was dissolved in mixture of ethyl acetate and water after evaporation of hydrogen fluoride, the free peptide was extracted into water and lyophilized. In the preparation of VI in larger scale, the protected peptide was treated with HF for 30 min at 0°C and after evaporation of HF purified by gel filtration (Sephadex G-15, 3M acetic acid). The yield was 51.2 mg (71%).

The HPLC Purification

The free cyclic peptide was dissolved in 5 ml 1M acetic acid and introduced onto the column containing Kromosil C18. The product was eluted by gradient running from 0% to 30% of acetonitrile in 60–120 min (see Table III), continuing isocratically at 30% of acetonitrile at the flow 60 ml/min. In the case of diastereoisomeric analogs prepared with D,L-Phe(*p*-Et) two peaks were isolated. The appropriate fractions were collected and lyophilized. Yields, HPLC constants, amino acid analyses and FAB MS are given in Table III.

Pharmacological Methods

Uterotonic activity *in vitro* was determined on an isolated strip of rat uterus^{23,24}. For determination of the uterotonic activity *in vivo* estrogenized rats under ethanol or Inactin anaesthesia were used²⁵. Pressor activity was determined on pithed rats²⁶. Inhibitory potencies were expressed by the pA_2 values^{27,28}. Prolongation of the uterotonic inhibitory activity was determined in rats in natural oestrus anaesthetized with Inactin⁹. The value of the inhibitor effect duration was determined as the time during which 75% of the original activity of the agonist (oxytocin) was retained after an inhibition of more than 60% of a standard dose of oxytocin.

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